

The Official Publication of the Canadian Academy of Audiology

Unraveling the Mystery of Hair Cell Death from Noise

Published July 9th, 2015

Eric Bielefeld, PhD
Deanna Meinke, Ph.D., CCC-A

Ever since a physician named Bernadino Ramazini (1700) first described tinnitus and hearing loss in coppersmiths in *De Morbis Artificum Diatriba* (*Diseases of Workers*), scientists have been trying to understand how noise damages the ear. At that time, Ramazini stated;

"To begin with, the ears are injured by that perpetual din, and in fact the whole head, inevitably, so that workers of this class become hard of hearing and, if they grow old at this work, completely deaf. For that incessant noise beating on the eardrum makes it lose its natural tonus; the air within

the ear reverberates against its sides, and this weakens and impairs all the apparatus of hearing." ¹

Over the next 150 years, scientists explored the gross anatomy of human cadavers to begin to explore the link between high levels of noise and hearing loss. Thurston provides an excellent summary of the general history of noise-induced hearing loss (NIHL) for those interested in the subject.²

With regard to the anatomical and physiological basis of NIHL, Fosbroke in 1831 appears to be the first to recognize that continuous noise exposure over a period of years results in *gradual onset*

hearing loss by the age of 40 or 50 years.³ In fact, his analogy regarding the cumulative nature of the noise damage from 1831 is just as relevant today as it was then;

"It is cumulative which makes it more difficult to guard against. It is like tobacco. One cigarette will not kill you and one day at work will not make you deaf. Neither will two or three – but where is the line? It is easy to say "one more exposure is not going to make any difference."

Fosbroke also stated that unidentified writers of that era attributed the gradual onset noise-induced hearing loss (NIHL) to paralysis of the auditory nerve, due to permanent over-tension of the

tympanic membrane.³ Toynbee differentiated sudden onset acoustic trauma from gunshots due to ankylosis of the stapes footplate as a different form of damage from exposure to continuous high

level noise sources attributed to expansion of the auditory nerve in the labyrinth.⁴ In 1882, as microscope technology and cell staining advancements were made, Habermann described atrophy to nerve fibers and the organ of Corti as a consequence of long-term noise exposure in a metal

worker that died suddenly after being struck by a locomotive he did not hear approaching.⁵ Hair cells, along with their specific locations, were not implicated until 1717 when Fraser and Fraser reported in the "The Morbid Anatomy of War Injuries of the Ear;

"(ii) The neuro-epithelium (hair cells) of Corti's organ are first affected, later the supporting cells are involved. The ganglion cells and nerve fibres are secondarily affected. The condition is one of so-called "degenerative neuritis." (iii) The part of the Corti's organ affected depends on the pitch

of the sound. If the noise be of high pitch the neuro-epithelim at the base of the cochlear is involved. If the noise be of medium pitch, Corti's organ in the middle coil is affected; while if the noise be of low pitch, degeneration is found in a portion of the Corti's organ nearer the apex."

Hallowell Davis' classic experiments in the 1940s further linked the hair cell damage to hearing loss and he also reported cochlear neuron degeneration and capillary vasoconstriction as a

consequence of hazardous noise exposure. Continued advancements in electron microscopes and cell staining techniques assist today's scientists in capturing and describing the histopathology of specific sound sources and exposure durations. We've known for almost 300 years that noise damages hair cells, it is the "HOW" of this damage that continues to challenge today's scientists such as Eric Bielefeld. What follows is Eric's personal story, and his attempt to "unravel" the complexities of cochlear cell death in the cochlea.

At the AudiologyNOW conference in 2002 in Philadelphia, I had the opportunity as a graduate student to help develop and present a talk on the mechanisms of hair cell injury underlying NIHL along with my fellow PhD student, Kelly Harris and my mentor, Don Henderson. In 2006, we expanded the presentation into a review paper for *Ear and Hearing* entitled, "Oxidative Stress in Noise-induced Hearing Loss." I believe that I learned more working on that article than I have for any other project in my career. Nearly 13 years after the presentation at AudiologyNOW, I had the opportunity to join Deanna Meinke at the 2015 National Hearing Conservation Association (NHCA) conference for another presentation on the mechanisms of hair cell death in noise-induced hearing loss. It was an opportunity to re-visit the topics from 2002 and 2006, review what has changed, and what still needs exploration and clarification. One thing is certain, the hair cell death process continues to become more complex as we learn more.

The first key topic underlying cochlear cell death from noise is oxidative stress. Oxidative stress results from an imbalance between reactive oxygen species (ROS) and antioxidants. Extensive evidence dating back to the early 1990s and continuing through current experimentation^{7–20} has implicated oxidative stress as a key underlying mechanism of NIHL. What remains largely speculative is the origin of the cochlear oxidative stress. Noise induces a burst of ROS formation that causes oxidative stress. ^{21–24} But the question of what causes the burst of ROS remains unanswered.

Multiple possibilities exist, and it is quite possible that multiple sources of ROS exist within the noise-exposed cochlea. Blood flow changes that result in ischemia (restriction of blood flow to tissue) and reperfusion (blood flow return to tissue after ischemia) are known to induce ROS formation (reviewed in Chan),²⁵ but the literature on the influence of noise on cochlear blood flow is inconsistent due to the use of different species, measurement times and approaches, and noise parameters.^{26–30} Therefore, cochlear blood flow changes may be a factor in ROS formation, but it is unclear how much of a factor.

Enzymatic reactions may also play a role in ROS formulation, specifically, the reaction in which the molecule NADPH donates an electron to molecular oxygen (O_2) to form superoxide (O_2) . This reaction is catalyzed by NDAPH oxidase. NADPH oxidase has been shown to be activated in the noise-exposed cochlea,³¹ and inhibition of NADPH oxidase in the cochlea reduces noise-induced threshold shift.³²

The severe metabolic demand placed on the cells of the cochlea that is exposed to persistent high-level noise has also been proposed as a source of ROS formation33 but the hypothesis has been

refuted on the grounds that over-driving of the mitochondria would deplete oxygen and ROS rather than producing them.³⁴ Instead it was proposed that dis-regulation of an enzyme in the Krebs' cycle of mitochondrial respiration, ?-ketoglutarate dehydrogenase, may be a key underlying event in cochlear ROS formation.³⁴ The cause of the dis-regulation of ?-ketoglutarate dehydrogenase in the noise-exposed cochlear cells remains the subject of hypotheses, and is a promising area for future investigation.

The second key topic to review about NIHL is the pattern of cell death. Cochlear cell death from noise is a primarily a combination of necrosis and apoptosis ^{9,33} along with an additional third pathway that does not fit either category's criteria. ³⁵ Necrosis is a passive form of cell death in which the cell and nucleus swell and eventually can burst open. Necrosis is not an energy-consuming process and can result in inflammation to tissue. Apoptosis is a controlled disassembly of the cell through a series of enzymatic reactions that initiate and execute the cell death process. Apoptosis is energy consumptive, and does not result in inflammation of the neighboring tissue.

The discovery of apoptosis in the noise-exposed cochlea was a major advancement in the understanding of NIHL, and for developing novel routes for prevention and treatment. What remains an enormous research undertaking is determining the sequence of molecular signals bridging from the noise insult to the complete execution of apoptotic cell death. Review of the literature reveals this cataloging to be an immense task for which considerable work still needs to be done. Since noise causes a combination of mechanical injuries to the structure of the organ of Corti and its individual cells, along with metabolic stress to the cells themselves, determining what stresses are triggering apoptosis is extremely challenging. To confuse the issue further, different noise exposures (for example long-duration continuous noise versus short-duration impulse noise) are likely to affect the organ of Corti in different ways and lead to different sequences of apoptosis.

Rather than beginning with the triggering event for apoptosis, it is clearer to begin with the execution of the cell's death and work backward toward the triggering event that initiates the sequence of events leading to apoptosis. For many routes of apoptosis, the final execution of cell death requires caspases. Caspase are enzymes that can initiate the apoptosis phase, or physically cleave the cell into pieces for export from the body. In particular, caspase-3 is an executor caspase whose action is one of the final events in many cells' apoptosis pathways. Caspase-3 activation has

been detected in the noise-exposed organ of Corti. Additionally, caspase-8 and caspase-9 were also detected. This is interesting because caspase-8 is heavily involved in the extrinsic apoptosis pathway, in which the toxic insult to the cell come from outside the cell and reaches the plasma membrane. Caspase-9 is involved in the intrinsic pathway, in which the apoptosis trigger originates from inside the cell at the mitochondria or nucleus. Seeing activation of both caspase-8 and -9 indicates multiple apoptosis sequences taking place concurrently within the same tissue.

Cytochrome c release from the mitochondria³⁶ and translocation of Endonuclease G from the mitochondria to the nucleus³⁷ have been detected in the noise-exposed cochlea. Both are key steps in the mitochondrial intrinsic apoptosis pathway, supporting its involvement in NIHL. Further implicating the mitochondrial pathway is the involvement of the Bcl-2 family, a complex family of proteins involved in intrinsic mitochondrial apoptosis, some of which signal for cell survival and some of which signal for cell death. The Bcl-2 cell survival proteins have been detected in the cochlea after noise exposure that induces temporary threshold shift (and thus the proteins promote recovery) while the pro-cell death Bcl-2 proteins have been detected after noise that induces permanent threshold shift.³⁸ While the mitochondrial intrinsic pathway is clearly involved in noise-

Canadian Audiologist - 3 / 5 - Printed 23.12.2025

induced apoptosis, it is certainly not the only major pathway, and the list of involved proteins provided here is not close to complete.

The next several years of research hold significant promise for further detailing the complex apoptosis signaling events in the noise-exposed cochlea, as well as identifying the triggering events the initiate the apoptosis process. As we learn more about the mechanisms of cochlear cell death, we will have the opportunity to intervene and prevent NIHL. The future will be exciting for audiology as we potentially become involved in administering antioxidant therapies in advance of noise exposure or preventing the cascade of events that lead to ROS formulation, cochlear cell death and NIHL post exposure.

References

- 1. Ramazzini B. 1964. Diseases of workers, translated from the Latin text de morbis artificum of 1713 by Wilmer Cave Wright. In: Wright CW, translator. New York: Hafner Publishing Company, 549 pp.
- 2. Thurston FE. The worker's ear: a history of noise-induced hearing loss. Am J Ind Med 2013;56(3):367–77.
- 3. Fosbroke J. 1831. Practical observations on the pathology and treatment of deafness. Lancet 1831:1:740–43.
- 4. Toynbee J. The diseases of the ear: Their nature, diagnosis, and treatment. Philadelphia: Blanchard and Lea; 1860.
- 5. Habermann J. Ueber die schwerho rigkeit der kesselschmied [On boilermaker's deafness]. Archiv fu r Ohrenheilkunde 1890;30(30):1–25.
- 6. Davis H. Temporary deafness following exposure to loud tones and noise. Acta Otolaryngol Suppl 1950;88.
- 7. Seidman MD, Shivapuja, BG, Quirk WS. The protective effects of allopurinol and superoxide dismutase on noise-induced cochlear damage. Otolaryngol Head Neck Surg 1993;109:1052–6.
- 8. Quirk WS, Shivapuja BG, Schwimmer CL, Seidman MD. Lipid peroxidation inhibitor attenuates noise-induced temporary threshold shifts. Hear Res 1994;74:217–20.
- 9. Hu BH, Henderson D, Nicotera TM. Involvement of apoptosis in progression of cochlear lesion following exposure to intense noise. Hear Res 2002;166:62–71.
- 10. Yamasoba T, Schacht J, Shoji F, Miller JM. Attenuation of cochlear damage from noise trauma by an iron chelator, a free radical scavenger and glial cell line-derived neurotrophic factor in vivo. Brain Res 1999;815:317–25.
- 11. Kopke RD, Weisskopf PA, Boone JL, et al. Reduction of noise-induced hearing loss using L-NAC and salicylate in the chinchilla. Hear Res 2000;149:138–46.
- 12. Kopke RD, Coleman JK, Liu J, et al. Candidate's thesis: enhancing intrinsic cochlear stress defenses to reduce noise-induced hearing loss. Laryngoscope 2002;112:1515–32.
- 13. Pourbakht A, Yamasoba T. Ebselen attenuates cochlear damage caused by acoustic trauma. Hear Res 2003;181:100–8.
- 14. Lynch ED, Gu R, Pierce C, Kil J. Ebselen-mediated protection from single and repeated noise exposure in rat. Laryngoscope 2004;114:333–7.
- 15. Bielefeld EC, Hynes S, Pryznosch D, et al. A comparison of the protective effects of systemic administration of a pro-glutathione drug and a Src-PTK inhibitor. Noise Health 2005;7:24–30.
- 16. Campbell KC, Meech RP, Klemens JJ, et al. Prevention of noise- and drug-induced hearing loss with D-methionine. Hear Res 2007;226:92–103.
- 17. Bielefeld EC, Kopke RD, Jackson RL, et al. Noise protection with N-acetyl-l-cysteine (NAC) using a variety of noise exposures, NAC doses, and routes of administration. Acta Otolaryng 2007;127:914–19.

- 18. Campbell K, Claussen A, Meech R, et al. D-methionine (D-met) significantly rescues noise-induced hearing loss: timing studies. Hear Res 2011;282:138–44.
- 19. Bielefeld EC, Wantuck R, Henderson, D. Post-exposure treatment with a Src-PTK inhibitor in combination with NAC to reduce noise-induced hearing loss. Noise Health 2011;13(53):292–8.
- 20. Claussen AD, Fox DJ, Yu XC, et al. D-methionine pre-loading reduces both noise-induced permanent threshold shift and outer hair cell loss in the chinchilla. Int J Audiol 2013;52:801–7.
- 21. Yamane H, Nakai Y, Takayama M, et al. The emergence of free radicals after acoustic trauma and strial blood flow. Acta Otolaryngol Suppl 1995;519:87–92.
- 22. Ohlemiller KK, Wright JS, Dugan LL. Early elevation of cochlear reactive oxygen species following noise exposure. Audiol Neurootol 1999;4:229–36.
- 23. Ohinata Y, Miller JM, Altschuler RA, Schacht J. Intense noise induces formation of vasoactive lipid peroxidation products in the cochlea. Brain Res 2000;878:163–73.
- 24. Yamashita D, Jiang HY, Schacht J, Miller JM. Delayed production of free radicals following noise exposure. Brain Res 2004;1019:201–9.
- 25. Chan PH. Reactive oxygen radicals in signaling and damage in the ischemic brain. J Cereb Blood Flow Metab 2001;2–14.
- 26. Perlman H, Kimura R. Cochlear blood flow in acoustic trauma. Acta Otolaryngol 1962;54:99–110.
- 27. Prazma J, Vance SG, Bolster DE, et al. Cochlear blood flow. The effect of noise at 60 minutes' exposure. Arch Otolaryngol Head Neck Surg 1987;113:36–9.
- 28. Thorne PR, Nuttall AL. Laser Doppler measurements of cochlear blood flow during loud sound exposure in the guinea pig. Hear Res 1987;27:1–10.
- 29. Ohlemiller KK, Dugan LL. Elevation of reactive oxygen species following ischemia-reperfusion in mouse cochlea observed in vivo. Audiol Neurootol 1999;4:219–28.
- 30. Lamm K, Arnold W. The effect of blood flow promoting drugs on cochlear blood flow, perilymphatic pO2 and auditory function in the normal and noise-damaged hypoxic and ischemic guinea pig inner ear. Hear Res 2000;141:199–219.
- 31. Ramkumar V, Whitworth CA, Pingle SC, et al. Noise induces A1 adenosine receptor expression in the chinchilla cochlea. Hear Res 2004;188:47–56.
- 32. Bielefeld EC. Reduction in noise-induced hearing loss with intra-cochlear application of an NADPH oxidase inhibitor. JAAA 2013;24:1–13.
- 33. Henderson D, Bielefeld EC, Harris KC, et al. The role of oxidative stress in noise-induced hearing loss. Ear Hear 2006;27:1–19.
- 34. Böttger EC, Schacht J. The mitochondrion: a perpetrator of acquired hearing loss. Hear Res 2013;303:12–9.
- 35. Bohne BA, Harding GW, Lee SC. Death pathways in noise-damaged outer hair cells. Hear Res 2007;223:61–70.
- 36. Nicotera TM, Hu BH, Henderson D. The caspase pathway in noise-induced apoptosis of the chinchilla cochlea. JARO 2003;4:466–77.
- 37. Zheng HW, Chen J, Sha SH. Receptor-interacting protein kinases modulate noise-induced sensory hair cell death. Cell Death Dis 2014;5:e1262.
- 38. Yamashita D, Minami SB, Kanzaki S, et al. Bcl-2 genes regulate noise-induced hearing loss. J Neurosci Res 2008;86:920–8.

Canadian Audiologist - 5 / 5 - Printed 23.12.2025